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# Review

# FIBRE EFFECTS ON NUTRITION AND REPRODUCTION IN PIGS

### Olusegun Matthew ATTEH\*, David Friday APATA

Department of Animal Production, Fisheries and Aquaculture, Kwara State University, Malete, P.M.B. 1530, Ilorin, Nigeria

# ABSTRACT

Dietary fibre (DF) is an indigestible carbohydrate in the form of a non-starch polysaccharide that is a major problem in the use of inexpensive agro-wastes by pigs such as brewer's dry grains, cassava peel, palm kernel cake, rice bran, sorghum spent grain and wheat offal. The DF in a pig's diet can be classified by its solubility as soluble dietary fibre and insoluble dietary fibre, and chemically – as neutral detergent fibre and acid detergent fibre. High DF levels reduced nutrient utilization, growth rate, in terms of pork production and reproductive performance. Pigs' reproductive cycles at different stages and lactation were both greatly impacted by increased DF intake. High DF decreased the number of stillborn piglets at birth and the farrowing duration by sow, while improving colostrum intake in piglets and colostrum yield in sows.

On the other hand, reducing the quantity of DF's hemicellulose in a sow's lactation diet can improve the low-birth-weight piglets' health, post-weaning performance and milk composition. While exogenous enzymes and the extrusion of fibrous feedstuffs enhanced nutrient digestibility and utilisation, other processing methods, including fermentation and extrusion, decreased the DF components and had a beneficial impact on the performance of the pigs. Agro-industrial residues have enormous potential to improve reproductive efficiency and serving as an economically viable DF feeding strategy in the pig industry.

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Key words: dietary fibre; pigs; performance; reproduction; blood parameters

# INTRODUCTION

The dietary fibre (DF) is an indigestible portion of the feed by pigs, chickens and humans and exerts vital impacts on maintaining normal physiological function and energy metabolism of the animal (Cummings and Stephen, 2007). DF are usually present in unconventional feedstuffs that are mostly agricultural by-products or industrial agro-waste, which are readily available and cheap for pigs feeding. Various plants and agricultural by-products, such as sweet potato peels, yam peels, cassava leaves, rice bran (RB), corn offal, cassava leaf meal, soy bean waste (SBW), sorghum spent grains (SSG), brewer's dry grain (BDG), corn bran, wheat offal and other locally available by-products, are characterized by high fibre content, low nutritional value and used in almost all households for feeding pigs in the developing world.

Agro-wastes in the environment are alternative feed resource usually used in feeding of monogastric animals as a result of continued rise in prices of conventional feedstuffs, such as maize and soya meal (Huang *et al.*, 2021), which are mostly used as sources of dietary energy and protein. Most agro-industrial by products, used in monogastric animal nutrition, contains non-starch polysaccharides (NSPs) in their cell wall that hindered digestibility by endogenous enzymes (Dalolio *et al.*, 2016). Low intake of DF by sows has negative effects on reproductive performance, increased stereotypic behaviour (Ramonet *et al.*, 1999), overweight of pregnant sows resulting in prolonged birth and giving birth to stillborn piglets.

**Copyright:** © 2024 Atteh & Apata **\*Correspondence:** E-mail: oluwaatteh63@gmail.com Olusegun Matthew Atteh, Department of Animal Production, Fisheries and Aquaculture, Kwara State University, Malete, P.M.B. 1530, Ilorin, Nigeria Received: October 1, 2023 Accepted: May 7, 2024

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The gastrointestinal tract of pig has the ability to utilise fibrous feeds of agro-industrial by-products by converting them into animal protein (Tonukari et al., 2016). High fibrous diets increased weight gain (Bertram et al., 2009; Gerritsen et al., 2012), while reduced growth or had no influence on growth performance accompanied by reduced digestibility of nutrients and energy (Jaworski et al., 2017). A higher proportion of crude fibre (CF) in the diet is linked to higher costs, a higher amount of feces and a higher amount of phosphorus excreted (Mpendulo et al., 2018). The fermentation of fibre results in small-chain fatty acids (SCFA) which enhances mucosal epithelial proliferation and villus height (Maswanganye et al., 2021). Nonetheless, supplementing DF to the diet in moderation may result in an increase in gut size, volume, length and morphological structure of pigs. Reproductive efficiency is the major factor that ascertain the sustainability of the pig industry. Fibre content in the diet has been shown to positively influence the reproductive efficiency of sows (Oliviero et al., 2019) through its physical features and influence on the development of the digestive tract (Wenk, 2001) as well as through the reproductive performance and the birth behaviour (Edwards et al., 2019). In sow, inclusion of DF in period prior to farrowing prevents constipation, increases water intake and feed intake during lactation, improves the duration of farrowing as well as milk intake and performance of piglets (Oliviero et al., 2019). This review focused on recent researches on utilisation of DF by swine at all stages of production and the consequences on their nutrition, and reproductive performance. Furthermore, help pig farmers understand alternative feed ingredients of low cost that are critical in reducing the dependence on conventional feed ingredients.

### **DIETARY FIBRE**

The DF is the major component of fiber-rich feed, accounting for about more than 40% of the total dry matter (DM) (Woyengo *et al.*, 2014). Fibre is defined as all carbohydrates that resist digestion in the upper gut of pigs. Carbohydrates of plants are divided into storage carbohydrates or starch and structural carbohydrates or non-starch polysaccharides (NSP). NSP can be grouped into soluble dietary fibre (SDF) and insoluble dietary fibre (IDF) based on their solubility

in water. SDF includes pectins, gums, and mucilages (Dhingra *et al.*, 2011).

Utilisation of fibrous feed ingredients by swine depends upon nature of the NSP. The main source of energy in pig diet is starch and digestion in the upper gut produces glucose. Source of DF in pigs include NSP which resist digestion in the upper gut and subsequently subjected to fermentation by gut microflora in the hindgut to produce volatile fatty acids (VFA) (Kerr and Shurson, 2013). DF can be fermented by intestinal microbes and provides 5 % – 28 % energy for pigs (Kass *et al.*, 1980).

Similarly, Yang and Zhao (2021), defined DF as neutral detergent fibre (NDF) and acid detergent fibre (ADF), which are commonly used in pig nutrition due to high cost of SDF and IDF analysis. The authors however, noted that NDF and ADF primarily contain cellulose, lignin, and insoluble hemicellulose, but SDF components are not measured. NDF and ADF are generally used analytical methods for the estimation of the nutritive qualities of feed. They were originally proposed by Van Soest and Wine (1968), but have undergone significant changes over the years (Mertens, 2003).

NDF was designed to isolate IDF components of plant cell walls like cellulose, hemicellulose, and lignin, whereas ADF is used to quantify cellulose and lignin (Dhingra *et al.*, 2011). SDF and IDF are dietary components usually recommended in application of pig production in order to achieve accurate feeding rather than ADF and NDF. Knudsen (2001) also recommended the use of IDF and SDF based on the physical properties of solubility in evaluating roles of DF in regulating pig nutrition accurately. SDF can be determined using high-performance liquid chromatography, or gravimetrically with ash and protein corrections (McCleary *et al.*, 2010).

Total dietary fibre (TDF) in feed ingredient contains IDF as the major component and SDF as a minor component (Jaworski *et al.*, 2015). The TDF, SDF and IDF can be determined using by the methods of Asp *et al.* (1983) with the use of the FIBERTEC machine by Tecator. Alternatively, SDF can be calculated as follows: organic matter – (protein + fat + soluble sugars + starch + NDF).

### Sources of dietary fibre in pigs' diets

Several agro-industrial wastes have been used in formulating diets for pigs especially in the tropics. Examples of non-conventional feed ingredients commonly used in swine nutrition; readily available and are relatively cheap are presented in Table 1. Corn bran consists of insoluble fibre with 280 g.kg<sup>-1</sup> cellulose and 700 g.kg<sup>-1</sup> hemicellulose comprising of bioactive components like corn fibre gum, cellulosic fibre gel, and xylo-oligosaccharides (Rose *et al.*, 2010). By-products from cassava tuber processing are usually high in starch (Régnier *et al.* 2010) and low in protein (Oguntimein, 1992) and, which are often use in replacement of maize at a least cost feed formulation.

The chemical and physical characteristics of DF can determine the rate of fermentation and VFA absorption. The negative impact of DF can vary considerably between fibre sources (Dunmire *et al.*, 2018). The crude protein (CP), ether extract (EE), CF, ADF, NDF, SDF and IDF of fibrous feedstuff were shown in Table 1. The CP value of 37.90 % of Dried brewers' grains (DBG) was shown to be the highest whereas, maize cobs (2.02 %) which is quite low in CP and highest in CF (28.69 %), ADF (51.58 %), NDF (70.63 %) when compared to other non-convention feed ingredients suggesting maize cob as a potential source of DF in

pig's nutrition. Szyszkowska *et al.*, (2007) observed that nutrient profile of maize cobs is affected by stage of maturity, cultivar, climate, soils and production methods. Mature cobs have higher NDF, ADF and lower CP than less mature cobs.

Fermentation of feeds have been a useful tool in improving the nutritive qualities of feeds for monogastrics (Apata and Atteh, 2016; Aladi, 2016). The microbial fermented feeds contains useful bacteria and animal digestive enzymes, helping the balance of intestinal flora and better digestion of nutrients (Le et al., 2016). Fermentation of feed components resulted in an increase in crude protein while decreasing the amount of CF, ADF, and NDF in fibrous feedstuffs, as shown in Table 1. The decreased in the various fibre components of fermented ingredients are due to the activities of enzymes released by microbes during the process of fermenting the feedstuff that brought about degradation of the non-cellulosic wall polysaccharides. The CP, SDF and IDF of corn bran fermented increased by 69.36 %, 53 % and 23 % respectively.

Ingredients	CP (%)	EE (%)	CF (%)	ADF	NDF	SDF	IDF
BDG <sup>a</sup>	25.04	9.02	18.00	21.00	37.55		
Corn offal <sup>ь</sup>	11.16	6.50	8.76	28.50			
CPM <sup>c</sup>	4.20	1.40	12.70	15.20	48.10		
CPLM <sup>d</sup>	18.93	2.67	16.78	32.97	43.28		
CW <sup>e</sup>	3.40	0.40	18.40	24.40	41.70		
CWF <sup>e</sup>	11.10	0.70	13.70	22.70	39.20		
DBG <sup>f</sup>	37.90	8.10		22.10	51.70		
DDGS <sup>g</sup>	31.10	7.90	8.50	12.00	27.90		
Maize cobs <sup>h</sup>	2.02	0.80	28.69	51.58	70.63		
Maize offal <sup>i</sup>	11.16	6.50	8.76	28.50			
PKC <sup>j</sup>	15.56	3.90		24.60	44.80	3.90	46.20
PKC <sup>k</sup>	17.50	10.70	16.80	46.80	12.40	79.00	45.90
PKC <sup>k</sup>	24.70	4.10	14.50	35.80	14.30	50.30	42.20
RB <sup>e</sup>	15.20	12.00	13.60	14.60	39.30		
RBF <sup>e</sup>	17.80	20.50	10.30	13.70	33.40		
SBP <sup>i</sup>	8.70	0.40	17.90	37.70	21.20	27.40	42.50
SSG <sup>a</sup>	25.59	8.00	14.00	21.00	42.88		
Wheat offal <sup>i</sup>	11.43	13.00	10.00				

#### Table 1. Chemical composition of fibrous feed ingredients in pig diet

CP: Crude protein; EE: Ether extract; CF: Crude fibre; ADF: acid detergent fibre; NDF: neutral detergent fibre; SDF: soluble dietary fibre; IDF: insoluble dietary fibre; BDG: brewer's dried grains; CPM: cassava peel meal; CPLM: cassava peel leaf meal; CW: cassava waste; CWF: cassava waste fermented; DBG: dried brewers' grains; DDGS: dried distillers grains with solubles; PKC: palm kernel cake; RB: rice bran; RBF: rice bran fermented; SBP: sugar beet pulp; SSG: sorghum spent grains "Babarinde *et al.* (2020); <sup>b</sup>Makinde and Sonaiya (2011); <sup>c</sup>Dayal *et al.* (2018); <sup>d</sup>Kehinde *et al.* (2020); <sup>e</sup>Hong *et al.* (2016); <sup>f</sup>\*Dunmire *et al.* (2018, 2010); <sup>j</sup>Olukayode *et al.* (2011); <sup>j</sup>Lv *et al.* (2022); <sup>k</sup>Marini *et al.* (2008); <sup>j</sup>Alikwe *et al.* (2012)

### UTILISATION OF DIETARY FIBRE BY PIGS

### Digestibility of fibre by pigs

Effects of fibre source on nutrient digestibility and fibre fermentability in pigs depends on the physiochemical properties of various fibre-rich ingredients (Mpendulo *et al.*, 2018), age of the animal as a result of differences in physiological stage of pigs. The digestibility of DF differs (40 % – 60 %) and very lower than other nutrients such as starch and sugars (above 80 %) (Berrocoso *et al.*, 2015), its digestibility was affected by the amount and the source of DF in the diet. DF in diets of pigs exhibited lower digestibility of dry matter, CP, gross energy, non-fibrous carbohydrates, and organic matter (Holt *et al.*, 2006; Feyera *et al.*, 2017). There is a high rate of degradation of DF in the hind gut of adult sows than in growing pigs due to longer retention time consecutive to their higher gastro intestinal tract volume combined with a lower feed intake per live weight (Jha and Berrocoso *et al.*, 2015).

According to studies by Kil *et al.* (2013) and Zhao *et al.* (2018), nutritional digestibility in growing pigs improves with age and breed and is largely affected by dietary management. The positive effect of increased fibre concentration on its digestibility is because more substrates went into the large intestine to be fermented.

Basal diets	Pig category/ phase	Inclusion level (%)	Effect	References
BDG	Weaner	30	Decreased performance	Imonikebe & Kperegbeyi (2014)
GSM	Piglets	15	Not significant ADFI, DWG & FCR	Martins <i>et al.</i> (2021)
SBP	Grower	15-55	Decreased apparent total	
			tract digestibility of dry matter,	
			crude protein and gross energy	Zhang <i>et al</i> . (2013)
Alfalfa meal	Weaner	60	Depressed daily gain and	
			efficiency of feed utilization	Kass <i>et al</i> . (1980)
Maize cobs	Finisher	7.5–15	Reduced weight gain	
			& feed intake	Frank <i>et al.</i> , (1983)
BSH	Finisher	58.50	Not significant	Olajide <i>et al.</i> (2019)
Maize cobs	Finisher	30	Reduced weight gain	
			& feed intake	Kanengoni <i>et al.</i> (2004)
BDG	Weaner	10	Improved performance	Amaefule et al. (2006)
DDGS	All pigs	30	Improved performance	Stein and Shurson (2009)
Maize bran	Growing	25	Improved performance	Akinfala <i>et al.,</i> (2017)
BNWM	Weaner pigs	40	Improved performance	Nwakpu <i>et al</i> .(2010)
FCSM	Weaners	6	Improved performance	Gu <i>et al</i> . (2021)
CP	Weaners	50	Improved performance	Oboh (2016)
РКС	Grower	30	Improved performance	Adesehinwa, 2007
CRP:PKC (3:1)	Weaner	55	Improved performance	
			and cost effective	Obongekpe (2020)
BDG	Weaners & growe	er 35	Improved performance at 40 %	
		and 40	weaners and 35 % for grower	Amaefule <i>et al.</i> (2006)
BDG	Grower	40	Improved digestibility	Amaefule <i>et al.</i> (2006)
CP + PKC	Grower	55	Improved performance	Oboh <i>et al.</i> (2018)
RB	Weaner	30	Improved performance	Adebiyi <i>et al.</i> 2018
RBF, CWF	Grower and finish	er 25	Reduced cost per kg,	
			reduced ADFI & ADWG	Hong <i>et al.</i> (2016)
DDGS	Finishing pigs		Poor feed to gain ratio	Dunmire <i>et al.</i> (2018)

### Table 2. Effects of dietary fibre on nutrition of pigs

ADFI: average daily feed intake; ADWG: average daily weight gain; BNWM: Bambara nut waste meal; BDG: Brewer's dry grains; BSH: CP: cassava peel; CRP: cassava root pulp; CWF: cassava waste fermented; DDGS: dried distillers grain with solubles; DWG: daily weight gain; FCSM: fermented cotton seed meal; FCR: feed conversion ratio; GSM: guava seed meal; PKC: Palm kernel cake; RB: rice bran; RBF: rice bran fermented; SBP: sugar beet pulp

Increasing dietary lipid content result to depressed fiber digestibility and more prominent at the faecal level than at the end of the ileum (Dégen et al., 2007). In a feeding trial by Wafar et al. (2020) different milling by-products using maize offal, wheat offal, sorghum offal, millet offal and rice offal respectively (Table 2). The result of growth performance and nutrient digestibility showed that average feed intake, final body weight as well as digestibility of dry matter, CF, EE, CP and ash were higher for pigs fed maize offal, wheat offal, sorghum offal and millet offal, while rice offal had the lowest growth and nutrient digestibility. It was concluded that maize offal, wheat offal, sorghum offal, millet offal can be used in weaned pigs' diet. Low nutrient digestibility in rice offal was attributed to the decrease in the mean retention time of this diet in the gastro-intestinal tract (Bindelle et al., 2008), depresses apparent digestibility of dry matter and nitrogen, decreases daily body weight and increase feed to gain ratio. The higher fibre digestibility observed for other cereal by-products were because of fermentation of DF in the hind gut with the VFA produced contributing the net energy requirement of the pigs. Yen et al. (1991) reported VFA contributing 5–28 % of energy required for maintenance in growing pigs resulted from fermentation of DF in the gastrointestinal.

Zhang *et al.* (2013) in their investigation observed that apparent total tract digestibility of dry matter, CP and gross energy decreased when TDF increased but the digestibility of SDF and IDF increased. The digestible energy (DE) and metabolisable energy (ME) content of diets decreased as the TDF increased and subsequently the amounts of nitrogen and energy excreted from feces were increased as DF level increased, while the amounts of energy excreted from urine was decreased. The DE and ME content of the diets decreased when the level of sugar beet pulp increased from 15.0 to 55.0 %.

# Performance and cost benefits of utilisation of fibre in pigs diet

Growing pigs with higher body weight have greater capacity to digest DF components than lower body weight of pigs. Many researchers have investigated the inclusion of DF at various growth stages of pigs (Table 2). Majority of research had focused on finding alternative to maize utilisation in the diets of pigs, leading to the use of fibrous feedstuff which tends to be cheaper and readily available. Le Goff and Noblet (2001) stated that greater capacity of heavy pigs and adult sows to digest DF is due primarily to more advanced development of pig intestine, rather than enhanced intrinsic ability of gut microbiota to degrade DF. Adesehinwa *et al.* (2017) conducted a feeding trial with weaned crossbred (Large white Landrace) pigs to determine the replacement value of cassava peel fine mash for maize. The result demonstrated the potential for replacement of up to 15 % of the 40 % maize inclusion in the diet.

Adesehinwa et al. (2011) reported that feeding an enzyme-supplemented cassava peel-based diet to growing pigs reduced the cost per kilogram of weight gain. Ekpo (2015) replaced maize with composite cassava tuber meal (C), BDG (B), and palm oil (P) mixed thoroughly at 65 %, 25 %, and 10 %, respectively, to produce CBP mixture in pig diets. The cost of producing a unit weight of pig was substantially reduced by replacing maize with CBP. CBP therefore may be utilised instead of maize in pig diets to reduce feed costs and total dependence on maize. Thus increase the farmer's profit when 50 kg of maize is replaced. Nwakpu et al. (2010) in an experiment using weaned piglets and recommend that it was cost effective in replacing 100 % maize with raw Bambara nut waste in the diets resulted to increased feed intake, increased daily weight gain when compared with the control diet and significantly reduced daily feed cost of feed per kg weight gain of pigs. Adesehinwa (2007) in replacing maize with palm kernel cake (PKC) on the performance and economy of production of growing pigs, concluded that maize when substituted with PKC (30 kg/100 kg of diet) improved performance characteristics and 50 % PKC replacement was the cheapest cost of production in diets of growing pigs. Adesehinwa et al. (2016) fed 30 % of cassava peel in replacement for 75 % of maize in growing pigs and discovered that there was no significant difference in body weight, feed intake and feed conversion ratio of between treatments and that it reduced 4 % in cost per kg of weight gain. They reported PKC as a high energy source and costeffective ingredient for ration formulation.

In a contrary view, Igene (2006) observed that feeding cassava peels to weaner pigs at level higher than 50 % decreased live weight and feed conversion efficiency. Similar observation was made by Amaefule *et al.*, (2006) replaced maize with BDG at 0, 30, 35 and 40 % BDG in pigs. Their results indicated that weaned pigs fed 40 % BDG diet had significantly higher protein intake (113.65 g) than others, while increasing levels of BDG in the diets significantly decreased feed cost at the weaned stage. And recommended 35 % of BDG at the grower stage for optimum performance. Frank *et al.* (1983) observed individual variability in ability of pigs to utilize the higher level of maize cob in their diets. The differences in responses to high fiber diets was attributed to genetic and physiological determinants of feed intake. Kanengoni *et al.* (2004) observed that in a growth performance experiment, growth rate decreased by 26 % and the Mukota pigs by 19 % as maize cobs increased from 0 to 300 g/kg in the diet.

Extrusion process inactivate the enzymes and subsequently prevent rancidity (Adebiyi et al., 2018). The temperature reached by the feed during extrusion cooking can be as high as 200 °C but the exposure time at this elevated temperature is very short (5 s to 10 s). Adebiyi et al. (2018) observed that extrusion processing of rice bran at 100 °C and 120 °C had a significant effect on weight gain and feed conversion ratio of weaner pigs when compared to those fed raw rice bran. This was attested to be as a result of the improvement in the crude protein, increased fibre digestibility. Adebiyi et al. (2018) concluded that extrusion treatment of rice bran increased the quality of rice bran for swine nutrition by feeding extruded rice bran at 120 °C up to 30 % inclusion improved weaner pigs' performance.

### Interaction of dietary fibre on intestinal mucosal of pigs

The gastrointestinal tract (GIT) help to digest and absorb nutrients from the diet. The GIT serves as a physical barrier that prevent the translocation of luminal toxins and antigens into epithelial immune cells. It also plays a major role in regulating epithelial and immune functions which are important for normal biological functioning and maintaining homeostasis in both the GIT and the body (Zhao et al., 2018). The GIT tract has its origin from the esophagus to the anus, composed of four concentric layers that constitute the wall of the tract: serosa, muscularis external, submucosa, muscularis mucosae and mucosa (Gartner and Hiatt, 2014). Intestinal mucin is the main constituent of the mucus protecting the gastrointestinal tract. For optimal mucosal protection, both the quantitative and qualitative characteristics of mucin are essential.

The intestinal mucosa is composed of epithelium, the gut-associated lymphoid tissue (GALT), and the mucus

overlying the epithelium. A delicate and dynamic equilibrium, which is crucial to a streamlined functioning and absorption capacity of the digestive system, is formed as a result of the interaction between the GALT, the intestinal mucus, host epithelial cells and microbiome (Maswanganye et al., 2021). Mucosa morphology reflects intestinal capacity of nutrient absorptions and digestion. Pigs hydrolyses and breaks down the ingredients of feed into tinier compounds; amino acids, peptides from proteins, fatty acids, monoglycerol from lipids and starch all contain glucose which is obtained by the mucosa. DF stimulate intestinal epithelial cells to secret mucosal protein and produce growth factors and metabolites such as arachidonic acid, all of which are beneficial to goblet cell proliferation and mucosal protein secretion (Yang and Zhao, 2021).

The fermentation of DF results in SCFA which enhances mucosal epithelial proliferation and villus height (Johnston et al., 2003). DF in wheat bran, pea fibre, cellulose have been shown to increase the excretion of intestinal mucin by stimulating the capacity of mucosal protein synthesis (Chen et al., 2013). Pea fibre (pea hulls and pea inner fibre) improved intestinal health in animals by reducing the adhesion and increasing the excretion of enterotoxigenic E. coli (Becker et al., 2009). A decreased ratio of villus height to crypt depth usually relates to impaired digestion and absorption of nutrients by intestinal mucosa. Yang and Zhao (2021) noted that DF directly disrupt surface structure of the mucosal layer resulting to increased speed of cell shedding, which causes compensatory growth of mucosal. And similarly, high inclusion of DF decreased energy density in diets, leading to an increase in feed intake and greater digesta flow in the intestine, which promotes renewal of mucosal protein, thus affecting intestinal mucosal layer. Addition of 10 % DF in the diets of growing pigs for a period of 14 days resulted in an increase in villi width and crypts depth in the ileum and jejunum which elevated rates of cell proliferation and crypt depth in the large intestines, as opposed to the same diet devoid of straw (Bach Knudsen, 2001).

Montagne *et al.* (2003) in their investigation observed that gut bacteria play a key role in interactive relationships between DF and mucosal layer or intestinal epithelium. But, DF have discrete structures, such as monosaccharide type, glycosidic bond, and physicochemical property, thus exerting different impacts on intestinal barrier in pigs. Nevertheless, moderate supplementation of DF to the diet resulted to an increase in gut size, volume, length and morphological structure of pigs (Maswanganye et al., 2021). Feeding pigs with high IDF diets might be better protected against pathogenic bacteria by increasing the villous length (Hedemann et al., 2006) and similar observation was made by Chen et al. (2013), who observed that DF elevated ileal mucosal integrity by improving ileal villous height and the villous height: crypt depth ratio. Feeding of pigs with IDF and SDF had more goblet cells in the ileum than those in a fibrefree group (Hino, S. et al., 2012). The mechanisms of the negative impact of high DF diets on nutrient digestibility could be as a result of soluble fibre that impair bile salt recycling and mixing and transport of micelles, which limits the effectiveness of the emulsification process and mass transfer of lipolytic products to the mucosal surface, thereby causing decreased nutrient digestion.

In addition, DF in pigs diets increase endogenous loss of nitrogen and amino acids due to enhanced secretion of digestive enzymes, mucin, and sloughed epithelial cells in the small intestine (Schulze *et al.*, 1994). Bikker *et al.* (2006) reported that feeding neonatal piglets high concentrations of SDF derived from wheat middlings, sunflower meal, and sugar beet pulp increased the length of the small intestine and improved amylase activity in the small intestinal brush border.

### Effect of dietary fibre on blood components of pigs

The haematology and serum biochemistry have been a useful assessment in predicting the nutritional condition of livestock. Several leaf meals of plant have been fed to pigs without being detrimental on blood parameters *Microdesmis puberula* leaves (Obua, 2013), cassava, Manihot (Iyayi, 2001), *Gmelina arborea* (Nkwocha *et al.*, 2008) and water hyacinth (Akovbovbo *et al.*, 2014). Olayemi *et al.*, (2006) opined that test ingredients have the ability to provide and maintain the essential amino acids and minerals in the diets which are necessary for the normal functioning of the blood cells producing tissues and organs.

Table 3 shows effects of feeding fibrous feedstuffs on blood parameters of pigs at different stages of their growth. High serum urea has been reported as an indicator of muscular wastage in animals (Adesehinwa, 2008). Ziemer *et al.* (2012) hypothesized that pigs feeding of fibrous feeds are likely to have high serum glucose concentrations resulting from increased availability of sugars from the fermentation of fibre. Nwakpu *et al.* (2010) fed raw bambara nut to weaned piglets in replacement for maize at 0, 10, 20, 30 and 40 and observed that all haematological indices had no negative effect on the animals (Table 3).

Obua (2013) fed Microdesmis puberula leaves that was harvested green, sun dried and ground into a meal and was subsequently fed to weaner pigs in replacement for soya bean meal. The study revealed that the RBC and Hb values of the pigs increased with increasing dietary inclusion of the leaf meal. The packed cell volume, haemoglobin and red blood cells were relatively high and best at 15 % dietary level showed enhanced quality of blood. Frank et al. (1983) reported a linear decrease in plasma glucose and an increase in plasma urea concentrations in cross-bred pigs fed diets with incremental levels of 7.5 % and 15 % maize cobs as fibre sources. The glucose decrease was attributed to less starch in the diet because maize cobs were incorporated in the diet at the expense of grain, while the high urea was because of increased ammonia production by intestinal microbes.

Blood urea is a principal product of the catabolism of protein and its concentration can act as an indicator of body protein status (Kohn et al., 2005). Serum energy metabolite concentrations depends on breed of pig and level of maize cobs in the same way their concentrations vary in response to type of fiber as reported by Weber and Kerr (2012). Mashatise et al. (2005) reported no differences in plasma glucose, urea and creatinine levels in Mukota and Mukota×Large White gilts fed a control and a high fiber diet with 20 % maize cobs. Adesehinwa et al. (2016) fed graded levels of high-quality cassava peel (HQCP) mash to growing pigs. Though the HDL-cholesterol and superoxide dismutase (SOD), was significantly different in the haematological and serum biochemical parameters among treatments, but all the values were within the normal range for healthy pigs and concluded that HQCP fine mash can effectively replace 75 % of the maize in the diet of growing pigs with no adverse effect on serum biochemical and haematological indices. The use of exogenous enzymes in fibrous feeds have been investigated by several authors on haematological and serum constituents of pigs. Adesinwa et al. (2008a) observed that 100 g Avizyme® 1300 inclusion in 100 kg of 45 %-CPM based diet as ideal for growing pig. Similarly, Adesehinwa et al. (2011) included Farmazyme which resulted in better utilisation of cassava peel meal. They observed that the protein levels of the maize-based diet and the CPM based diet supplemented with Farmazyme®

Feed ingredient	Pig category/ phase	Optimum inclusion level	Effect	References
BNWM	Weaner	40	Haematological data not significant	Nwakpu <i>et al.</i> (2010)
MPLM	Weaner	15	Normal haematology and serum biochemistry	Obua (2013)
IALM	Grower	15	Normal haematology and serum biochemistry	Ekenyem & Madubuike (2007)
СРМ	Grower	30	no significant effect on serum metabolites, haematological parameters and serum electrolyte but for slight variations observed in the values of MCHC and PO4 <sup>2-</sup>	Adesehinwa <i>et al.</i> (2008) es
PBSM	Weaner	11-15	Negative effect on haematology and serum components	Ogunbode <i>et al.</i> (2016)
СРМ	Grower	40	Normal haematology and serum biochemistry	Irekhore <i>et al.</i> (2015)
HQCP CRP:PKC (3:1)	Weaner	30 55	Haematology and serum were positively correlated	Adesehinwa <i>et al.</i> (2016) Obongekpe (2020)
Maize offals	weaners	26.45	Serum not significantly different from control	Adesehinwa <i>et al.</i> (1999)
TDLM	Growers	21-30	Negative effect on haematology and serum components	Fasuyi <i>et al.</i> (2013)

### Table 3. Effects of feeding dietary fibre on haematology and serum of pigs

BNWM: Bambara nut waste meal; CPM: Cassava peel meal; CRP: cassava root pulp; HQCP: high quality cassava peel; IALM: *Ipomoea asarifolia* leaf meal; MPLM: *Microdesmis puberula* leaf meal; PBSM: Pride of barbados seed meal; PKC: palm kernel cake; PBSM: pride of barbados seed meal; TDLM: *Tithonia diversifolia* (wild sunflower) leaf meal

was able to support the protein reserves of the pigs across the groups and that CPM was efficiently utilised leading to high tissue deposition.

# Effect of dietary fibre on reproductive performance of pigs

In large-scale pig production, sows and piglets are crucial to determining production levels and the economic benefits of pig farms, and the gestational, lactation, and newborn periods are critical stages for feed management of sows and piglets in large-scale pig production (Kim *et al.*, 2013). Feeding of fibrous feeds to pigs improved their reproductive performance at different stages. Proper feeding of pregnant sows improved feed intake which can help to prevent constipation thus enhancing their reproductive performance. Origin and type of DF determine the reproductive effects on the animals. DF improved oocyte quality and early embryo survival rate, increased litter size, prevent gestation and miscarriage, and improved the reproductive performance of sows (Loisel *et al.*, 2013; Yin *et al.*, 2016). DF reduced estrogen during follicular development that could improve the survival rate of early embryos and reduced intrauterine growth retardation of fetal pigs, even stimulated the immune system, and reduced diarrhea rate and mortality (Ferguson *et al.*, 2007). Feeding a high DF prior to mating also improved the outcome of pregnancy. Proper feeding of pregnant sows improved feed intake which can help to prevent constipation thus enhancing their reproductive performance.

The type of DF fed to pregnant sows helps to determine the reduction farrowing length and stillborn piglets. DF influence the energy metabolism of the sow (Le Goff and Noblet 2001), inclusion of exogenous enzymes helps to improve the energy digestibility of the diet (Zhou *et al.*, 2009) and consequently allow a coarser diet, which is central in order to avoid stomach

ulcers. High DF diets intake during the perinatal period were documented to reduced prolonged farrowing duration by softening the faeces and supplying energy from the hindgut (Loisel *et al.*, 2013; Feyera *et al.*, 2017). Meunier-Salaün *et al.* (2001) observed that energy requirement of sows' increases at last weeks of gestation which justifies reasons why the raw fibre requirement for gestating sows is reduced to a minimum. Feeding sows a diet high in fermentable NSP during the weaning to oestrus interval and the subsequent pregnancy over three successive parities, resulted in an increase in the total number of piglets born and the number of live born piglets (Van der Peet-schwering *et al.*, 2003).

Higher proportion of SDF to IDF in the pregnant diet of sows improved the antioxidant capacity and inhibited the inflammation of the colon for piglets (Li et al., 2020). SDF intake promote neonatal intestinal health through maternal inulin intake during late pregnancy and lactation changed the intestinal microbiota for their suckling piglets (Paßlack et al., 2015). Research on maternal SDF intake by pregnant sows needed to be explore because of its beneficial effect to the intestinal health of piglets. Collaborating this, Holt et al. (2006) observed that DF for gestating sows during transition reduced the proportion of stillborn piglets and mortality of total born piglets. According to the Animal Welfare and Animal Husbandry Ordinance, in Germany breeding sows must be fed with at least 8 % or 200 g CF per day per animal up until one week before the expected farrowing date (Ernährung, 2006).

Che et al. (2011) reported the effect of feeding DF on reproductive performance during the first two parities. The authors discovered that at first parities; sows fed low DF diet in parity 1 gained more backfat during gestation but lost more during lactation than sows fed high DF diet; sows fed low DF diet farrowed more pigs (+0.7-1.1 pigs) and pigs born alive (+1.0 pigs) relative to sows fed high DF diet. Similarly, sows fed Low DF diets had greater litter weights at parturition (both p = 0.06) and day 22 after lactation. In parity 2, after 22 d of lactation, sows fed high DF diet had most pigs alive and heavier litter weight, increased internal organs weight of newborns. Papatsiros et al. (2021) reported that feeding of sows with high DF lead to a decrease of the number of stillborn piglets at birth, increase of the number of live born and weaned piglets and reduced farrowing time. Darroch et al. (2008) added 20 % soybean hulls and 0.3 % psyllium to the diets of pregnant sows. Their results showed that soybean hull was more conducive to physical health maintenance in pregnant sows but had little effect on litter size.

Beneficial bacteria in the intestine with functional SDF help improve the metabolic syndrome in the way of "microbiota remodeling", thereby alleviating inflammation and oxidative stress in sows, which effectively increased the ADFI of sows during lactation (Tan et al., 2016; Li et al., 2020; Xu et al., 2020). Zhuo et al. (2020) found that IDF of oat bran mixed with corn or soybean meal produced more SCFA by gut fermentation, which improved pig behaviors and reproductive performance. Cheng et al. (2018) added combined SDF from pregelatinized waxy corn starch and guar gum to the sows' pregnancy diet, which significantly improved the developmental growth performance and gut function of 14-day-old suckling piglets. Research on effect of feeding DF during lactation on the performance of suckling piglets is scanty because of the ability of fibrous ingredients to reduce feed intake. The number of piglets weaned was affected by feeding DF during gestation.

Researchers have shown that the nutritional composition of colostrum can be changed with additives or types of DF in the diet of pregnant and lactating sows, thereby regulating the growth performance and immunity of offspring piglets (Grela et al., 2019; Tan et al., 2021; Zhang et al., 2020). DF reduced the concentration of dietary energy, but improved the colostrum intake by piglets and colostrum yield of gestating sows (Theil et al., 2014). Luo et al. (2021) fermented Radix puerariae residue (FRPR), their results showed that dietary FRPR had no effects on litter size and the number of total alive piglet, and that the number of weaned piglets and weaning weight of litter were increased in sows with 4 % FRPR treatment compared with control treatment. The addition of guar gum to maternal diets increased the population of beneficial microbiota in the intestine and reduced the diarrhea rate of piglets (Cheng et al., 2018).

# CONCLUSION

The current review demonstrated that feeding DF to pigs at all stages of growth, especially during the growing and finishing periods, is nutritionally beneficial. CPM, PKC, and RB improved the productive performance and economic efficiency of growing pigs; SDF in SBP improved enzyme activities in the small intestinal walls; fermentation of CW and rice bran improved crude protein while reducing ADF and NDF; enzyme supplementation resulted in high tissue deposition; extrusion of RB at 120 °C improved performance of weaned pigs, and DF reduced piglet stillbirth and mortality. Agro-industrial residues have major potential as economically viable DF feeding strategies in pig production, as well as for improving reproductive efficiency.

# AUTHOR'S CONTRIBUTIONS

Conceptualization: Atteh, O. M.

Methodology: Apata, D. F.

Investigation: Atteh, O. M.

Data curation: Apata, D. F.

Writing-original draft preparation: Atteh, O. M.

Writing-review and editing: Atteh, O. M.

Project administration: Apata, D. F.

All authors have read and agreed to the published version of the manuscript.

# DATA AVAILABILITY STATEMENT

The data presented in this study are available on request from the corresponding author.

# **CONFLICT OF INTEREST**

The authors declare no conflict of interest.

# REFERENCES

- Adebiyi, O. A., Sodeke, M., Adeleye, O. O. & Adejumo, I. O. (2018). Effects of extruded rice bran-based diets on the performance, intestinal microbiota and morphology of weaned pigs. *Agricultura Tropica et Subtropica*, 51(1), 13–19.
- Adesehinwa, A. O. K., Fanimo, A. O., Ogunsanmi, A., Uma, M. & Ogunmodede, B. K. (1999). Weaner pigs fed. graded levels of maize offal as replacement for maize. 2. Effect on serum metabolites. *Tropical Journal Animal Science*, 1(2), 35–43.
- Adesehinwa, A. O. K. (2007). Utilization of palm kernel cake as a replacement for maize in diets of growing pigs:

effects on performance, serum metabolites, nutrient digestibility and cost of feed conversion. *Bulgarian Journal of Agricultural Science*, 13, 593–600.

- Adesehinwa, A. O. K. (2008a). Energy and protein requirements of pigs and the utilization of fibrous feedstuffs in Nigeria. *African Journal of Biotechnology*, 7(25), 4798–4806.
- Adesehinwa, A. O. K., Dairo, F. A. S. & Olagbegi, B. S. (2008b). Response of growing pigs to cassava peel-based diets supplemented with Avizyme® 1300: growth, serum and hematological indices. *Bulgaria Journal of Agricultural Science*, 14, 491–499.
- Adesehinwa, A. O. K., Obi, O. O., Makanjuola, B. A., Oluwole, O. O. & Adesina, M. A. (2011). Growing pigs fed cassava peel based diet supplemented with or without Farmazyme 3000 proenx: Effect on growth, carcass and blood parameters. *African Journal of Biotechnology*, 10(14), 2791–2796.
- Adesehinwa, A. O. K., Samireddypalle, A., Fatufe, A. A., Ajayi, E., Boladuro, B. & Okike, I. (2016). High quality cassava peel fine mash as energy source for growing pigs: effects on performance, cost of production and blood parameters. *Livestock Research for Rural Development*, 28, 11, Art.No. 207.
- Adesehinwa, A. O. K., Fatufe, A. A., Samireddypalle, A., Ajayi, E., Adetunji, T. A., Okike, I. & Adesehinwa, O. (2017). Utilization of high quality cassava peel (HQCP<sup>®</sup>) mash as an alternative source of energy in weaned pigs diet. *Proceeding of the American Society of Animal Science Conference*, July 8–12, 2017, Baltimore Convention Center, Maryland, USA.
- Akinfala, E. O., Ogundeji, S. T. & Adewole, E. F. (2017). Growth response, carcass traits and cholesterol of growingfinishing pigs fed different fibre feedstuffs-based diets. *Nigerian Journal of Animal Science*, 2, 114–122.
- Akovbovbo, G., Ajuogu, P. K. & Yahaya, M. A. (2014). Haematological response of weaners pigs fed with graded levels of water hyacinths (*Eichornia crassipes*) as feed supplement. *International Journal Advanced Biological Research*, 4(2), 165–167.
- Aladi, N. O. (2016). *Studies on dietary fermented mixture of cassava and palm kernel cake on carcass characteristics of broilers and pigs*. Ph.D. Thesis, Federal University of Technology, Owerri, Nigeria.
- Alikwe, P. C. N., Lamidi, A. A. & Aina, A. B. J. (2012). Comparative digestibility and nitrogen balance of maize bran, wheat offal and rice bran in West African dwarf goats. *Journal* of Agriculture and Social Research, 12(1), 108–113.
- Amaefule, K. U., Okechukwu, S. O., Ukachukwu, S. N., Okoye, F. C. & Onwudike, O. C. (2006). Digestibility and nutrient utilization of pigs fed graded levels of

brewers' dried grain-based diets. *Livestock Research for Rural Development*, 18(1), Art.No. 5.

- Apata, D. F. & Atteh, O. M. (2016). Growth performance and intestinal morphology of broiler chickens fed diets containing almond fruit fermented with Aspergillus niger. Wayamba Journal of Animal Science, 1436–1444.
- Asp, N. G., Johansson, C. G., Hallmer, H. & Siljestrom, M. (1983). Rapid enzymatic assay of insoluble and soluble dietary fibre. *Journal of Agricultural Chemistry*, 31, 476–482.
- Bach Knudsen, K. E. (2001). The nutritional significance of "dietary fibre" analysis. *Animal Feed Hermes, Science and Technology*, 90(1), 3–20.
- Becker, P. M., Wikselaar, P. G. & Jansman, A. J. (2009). Pea dietary fibre for adhesion an excretion of enterotoxigenic *E. coli* K88 to prevent intestinal colonization. *Journal of Animal Science*, 87, 172–189.
- Berrocoso, J. D., Menoyo, D., Guzmán, P., Saldaña, B. & Cámara, L. & Mateos, G. G. (2015). Effects of fiber inclusion on growth performance and nutrient digestibility of piglets reared under optimal or poor hygienic conditions. *Journal of Animal Science*, 93(8), 3919–3931.
- Bertram, H. C., Malmendal, A., Nielsen, N. C., Straadt, I. K., Larsen, T., Knudsen, K. E. & Laerke, H. N. (2009). NMRbased metabolomics reveals that plasma betaine increases upon intake of high-fiber rye buns in hypercholesterolemic pigs. *Molecular Nutrition & Food Research*, 53(8), 1055–1062.
- Bikker, P., Dirkzwager, A., Fledderus, J., Trevisi, P., Le Huerou-Luron, I., Lalles, J. P. & Awati, A. (2006) The effect of dietary protein and fermentable carbohydrates levels on growth performance and intestinal characteristics in newly weaned piglets. *Journal of Animal Science*, 84, 3337–3345.
- Bindelle, J., Leterme, P. & Buldgen, A. (2008). Nutritional and environmental consequences of dietary fibre in pig nutrition: A review. *Biotechnolology Agronomy, Society and Environment*, 12, 69–80.
- Che, L., Feng, D., Wu, D., Fang, Z., Lin, Y. & Yan, T. (2011). Effect of dietary fibre on reproductive performance of sows during the first two parities. *Reproduction in Domestic Animals*, 46, 1061–1066. DOI:10.1111/ j.1439-0531.2011.01787.x
- Chen, H., Mao, X., He, J., Yu, B., Huang, Z., Yu, J., Zheng, P. & Chen, D. (2013). Dietary fibre affects intestinal mucosal barrier function and regulates intestinal bacteria in weaning piglets. *British Journal of Nutrition*, 110, 1837–1848.

- Cheng, C., Wei, H., Xu, C., Xie, X., Jiang, S. & Peng, J. (2018). Maternal soluble fiber diet during pregnancy changes the intestinal microbiota, improves growth performance, and reduces intestinal permeability in piglets. *Applied and Environmental Microbiolobiology*, 84(17), e01047–18.
- Cummings, J. H. & Stephen, A. M. (2007). Carbohydrate terminology and Classification. *European Journal of Clinical Nutrition*, 61(1), S5–S18.
- Dalolio, F. S., Moreira, J., Vaz, D. P., Albino, L. F. T., Valadares, L. R., Pires, A. V. & Pinheiro, S. R. F. (2016). Exogenous enzymes in diets for broilers. *Revista Brasileira de Saúde e Produção Animal*, 17, 149–161.
- Darroch, C. S., Dove, C. R., Maxwell, C. V., Johnson, Z. B. & Southern, L. L. S. (2008). Regional research committee on nutrition and management of swine for increased reproduction efficiency. A regional evaluation of the effect of fiber type in gestation diets on sow reproductive performance. *Journal of Animal Science*, 86, 1573–1578.
- Dayal, A. D., Diarra, S. S., Devi, S. L. A & Amosa, F. (2018). High cassava peel meal-based diets with animal fat and enzyme for broilers. *Livestock Research for Rural Development*, 30(6).
- Dégen, L., Halas, V. & Babinszky, L. (2007) Effect of dietary fibre on protein and fat digestibility and its consequences on diet formulation for growing and fattening pigs: A review. Acta Agriculturae Scandinavica, Section A – Animal Science, 57(1), 1–9.
- Dhingra, D., Michael, M. & Rajput, H. (2011). Dietary fibre in foods: A review. *Journal of Food Science and Technology*, 49, 255–266.
- Dunmire, K. M., Thomas, L. L., Braun, M. B., Truelock, C. N., Tokach, M. D., DeRouchey, J. M., Goodband, R. D., Woodworth, J. C., Dritz, S. S. & Paulk, C. B. (2018).
  Effect of dietary fiber source on growth performance, carcass characteristics, and economic return of finishing pigs. *Kansas Agricultural Experiment Station Research Reports*, 4(9). DOI: 10.4148/2378-5977.7680
- Edwards, L., Plush, K. J., Ralph, C. R., Morrison, R. S., Acharya, R. Y. & Doyle, R. E. (2019). Enrichment with Lucerne Hay Improves Sow Maternal Behaviour and Improves Piglet Survival. *Animals*, 9(8), 558.
- Ekenyem, B. U. & Madubuike, F. N. (2007). Haematology and serum biochemistry of grower pigs fed varying levels of *Ipomoea asarifolia* leaf meal. *Pakistan Journal of Nutrition*, 6(6), 603–606.
- Ekpo, J. S. (2015). Comparative evaluation of yam peel meal and pumpkin stem waste on the performance and carcass characteristics of growing pigs. *Russian*

Agricultural Sciences, 41, (4), 297–299.

- Fasuyi, A. O., Ibitayo, F. J. & Alo, S. O. (2013). Histopathology, haematology and serum chemistry of growing pigs fed varying levels of wild sunflower (*Tithonia diversifolia*) leaf meal as protein supplements. *IOSR Journal of Agriculture and Veterinary Science*, 4(1), 41–50.
- Ferguson, E. M., Slevin, J., Hunter, M. G., Edwards, S. A. & Ashworth, C. J. (2007). Beneficial effects of a high fibre diet on oocyte maturity and embryo survival in gilts. *Reproduction*, 133(2), 433–439. DOI: 10.1530/REP-06-0018
- Feyera, T., Hojgaard, C. K., Vinther, J., Bruun, T. S. & Theil, P. K. (2017). Dietary supplement rich in fiber fed to late gestating sows during transition reduces rate of stillborn piglets. *Journal of Animal Science*, 95(12), 5430–5438. DOI: 10.2527/jas2017.2110
- Frank, G. R., Aherne, F. X. & Jensen, A. H. (1983). A study of the relationship between performance and dietary component digestibilities by swine fed different levels of dietary fiber. *Journal of Animal Science*, 57, 645–654.
- Gartner, L. P. & Hiatt, J. L. (2014). *Color Atlas and Text of Histology*. (6<sup>th</sup> ed.). Baltimore: Lippincott Williams & Wilkins. ISBN 978-1-4511-1343-3.
- Gerritsen, R., van der Aar, P. & Molist, F. (2012). Insoluble nonstarch polysaccharides in diets for weaned piglets. *Journal of Animal Science*, 90, 318–320.
- Grela, E. R., Czech, A., Kiesz, M., Wlazło, Ł. & Nowakowicz-Debek, B. (2019). A fermented rapeseed meal additive: effects on production performance, nutrient digestibility, colostrum immunoglobulin content and microbial flora in sows. *Animal Nutrition*, 5, 373–379.
- Gu, X., Chen, J., Li, H., Song, Z., Chang, L., He, X. & Fan, Z. (2021). Isomaltooligosaccharide and *Bacillus* regulate the duration of farrowing and weaning-estrous interval in sows during the perinatal period by changing the gut microbiota of sows. *Animal Nutrition*, 7(1), 72–83.
- Hedemann, M. S., Eskildsen, M., Lærke, H. N., Pedersen, C., Lindberg, J. E., Laurinen, P. & Bach Knudsen, K. E. (2006). Intestinal morphology and enzymatic activity in newly weaned pigs fed contrasting fiber concentrations and fiber properties. *Journal of Animal Science*, 84(6), 1375–1386.
- Hino, S., Takemura, N., Sonoyama, K., Morita, A., Kawagishi, H., Aoe, S. & Morita, T. (2012). Small intestinal goblet cell proliferation induced by ingestion of soluble and insoluble dietary fiber is characterized by an increase in sialylated mucins in rats. *Journal of Nutrition*, 142(8), 1429–1436.
- Holt, J. P., Johnston, L. J., Baidoo, S. K. & Shurson, G. C.(2006). Effects of a highfiber diet and frequent feeding on behavior, reproductive performance, and nutrient

digestibility in gestating sows. *Journal of Animal Science*, 84, 946–9455.

- Hong, T. T. T., An, L. V., Be, P. T. & Lindberg, J. E. (2016). Effect of fermented rice bran and cassava waste on growth performance and meat quality of crossbred pigs. *World Journal of Agricultural Research*, 4(5), 132–138.
- Huang, B., Wang, L., Lyu, Z., Wang, L., Zang, J., Li, D. & Lai, C. (2021). Evaluation on net energy of defatted rice bran from different origins and processing technologies fed to growing pigs. *Animals*, 11(4), 1106.
- Igene., F. U. (2006). Essentials of Pigs Production in Nigeria J.L.G Publishers, Ibadan, 1<sup>st</sup> Edition.
- Imonikebe, U. G. & Kperegbeyi, J. I. (2014). Effect of substitution of maize with brewer's dried grain in pig starter diet on the performance of weaner pig. *Global Journal of Agricultural Research*, 2(4), 42–48.
- Irekhore, O. T., Adeyemi, O. M., Idowu, O. M. O., Akinola, O. S. & Bello, K. O. (2015). Growth performance, haematological indices and cost benefits of growing pigs fed cassava peel meal diets supplemented with Allzyme<sup>®</sup> SSF. International Journal of Applied Agricultural Research, 11, (1&2), 51–59.
- Iyayi, E. A. (2001). Cassava leaves as supplements for feeding weaner swine. *Tropical Animal Production Investment*, 4, 141–150.
- Jaworski, N. W. & Stein, H. H. (2017). Disappearance of nutrients and energy in the stomach and small intestine, cecum, and colon of pigs fed corn-soybean meal diets containing distillers dried grains with solubles, wheat middlings, or soybean hulls. *Journal of Animal Science*, 95(2), 727–739.
- Jaworski, N. W., Lærke, H. N., Bach Knudsen, K. E. & Stein, H. H. (2015). Carbohydrate composition and *in vitro* digestibility of dry matter and non-starch polysaccharides in corn, sorghum, and wheat and coproducts from these grains. *Journal of Animal Science*, 93, 1103–1113.
- Jha, R. & Berrocoso, J. D. (2015). Review: dietary fiber utilization and its effects on physiological functions and gut health of swine. *Animal*, 9, 1441–1452.
- Johnston, L. J., Noll, S., Renteria, A. & Shurson, J. (2003). Feeding by-products high in concentration of fiber to nonruminants. In: National Symposium on Alternative Feeds for Livestock and Poultry. Kansas City, MO, 169–186.
- Kanengoni, A. T., Dzama, K., Chimonyo, M., Kusina, J. & Maswaure, S. M. (2004). Growth performance and carcass traits of Large White, Mukota and Large White × Mukota F1 crosses given graded levels of maize cob meal. *Animal Science*, 78, 61–66.

- Kass, M. L., Van Soest, P. J., Pond, W. G, Lewis, B. & McDowell, R. E. (1980). Utilization of dietary fiber from alfalfa by growing swine. I. Apparent digestibility of diet component in specific segments of the gastrointestinal tract. *Journal of Animal Science*, 50, 175–191. DOI: 10.2527/jas1980.501175
- Kehinde, A. S., Babatunde, T. O., Kehinde, J. O., Babatunde, O. O., Adelakun, K. M. Fadimu, B. O., Abdulazeez, F. I.
  & Ogundimu, A. O. (2020). Fibre characterization of cassava peel leaf meal and its utilization by broilers. *Journal of Applied Science and Environmental Management*, 24(9), 1529–1533.
- Kerr, B. J. & Shurson, G. C. (2013). Strategies to improve fiber utilization in swine. *Journal of Animal Science* and Biotechnology, 4(1), 11.
- Kohn, R. A., Dinneen, M. M. & Russek-Cohen, E. (2005). Using blood urea nitrogen to predict nitrogen excretion and efficiency of nitrogen utilization in cattle, sheep, goats, horses, pigs, and rats. *Journal of Animal Science*, 83, 879–889.
- Kil, D. Y., Kim, B. G. & Stein, H. H. (2013). Feed energy evaluation for growing pigs. *Asian-Australis Journal of Animal Sciences*, 26(9), 1205–1217.
- Kim, S. W., Weaver, A. C., Bin Shen, Y. & Zhao, Y. (2013). Improving efficiency of sow productivity: Nutrition and health. *Journal of Animal Science Biotechnology*, 4(1), 26.
- Knudsen, K. E. B. (2001). The nutritional significance of "dietary fibre" analysis. *Animal Feed Science and Technology*, 90, 3–20.
- Le Golf, G. & Noblet, J. (2001). Comparative digestibility of dietary energy and nutrients in growing pigs and adult sows. *Journal of Animal Science*, 79, 2418–2427.
- Le, M. H. A., Galle, S., Yang, Y., Landero, J. L., Beltranena, E., Ganzle, M. G. & Zijlstra, R. T. (2016). Effects of feeding fermented wheat with *Lactobacillus reuteri* on gut morphology, intestinal fermentation, nutrient digestibility, and growth performance in weaned pigs. *Journal of Animal Science*, 94(11), 4677–4687.
- Li, H., Liu, Z., Lyu, H., Xueling, G., Song, Z., He., X., & Fan, Z. (2020). Effects of dietary inulin during late gestation on sow physiology, farrowing duration and piglet performance. *Animal Reproduction Science*, 106531.
- Loisel, F., Farmer, C., Ramaekers, P. & Quesnel, H. (2013). Effects of high fiber intake during late pregnancy on sow physiology, colostrum production, and piglet per formance. *Journal of Animal Science*, 91, 5269–5279. DOI: 10.2527/jas.2013-6526
- Luo, Z., Zhao, Y., Zeng, L., Yin, J., Zeng, Q., Li, X., He, J., Wang, J. & Tan, B. (2021). Effects of fermented radix

puerariae residue on nutrient digestibility and reproductive performance of sows. *Frontier Nutrition*, 8, 715713. DOI: 10.3389/fnut.2021.715713

- Lv, Z., Zhang, Z., Wang, F., Guo, J., Zhao, X. & Zhao, J. (2022). Effects of dietary fiber type on growth performance, serum parameters and fecal microbiota composition in weaned and growing-finishing pigs. *Animals*, 12(12), 1579.
- Makinde, O. A. & Sonaiya, E. B. (2011). Utilization of sun-dried maize offal with blood meal in diets for broiler chickens. *Open Journal of Animal Science*, 1(3), 106–111.
- Marini, A. M., Ayub, M. Y., Abd. Salam, B. Hadijah, H., Azahan, E. A. E. & Tarmizi, S. A. (2008). Protein quality of *Aspergillus niger*-fermented palm kernel cake. *Journal of Tropical Agriculture and Food Science*, 36(2), 227–237.
- Martins, J. S., Genova, J. L., Leal, I. F., Barbosa, K. A., Santos,
  L. B., Rupolo, P. B., Reis, L. E., Escocard de Oliveira, N.
  T., Carvalho, P. O. & Bruno, L. D. G. (2021). Potential impacts of guava seed meal on piglet feeding as a dietary fibre alternative. *Journal of Applied Animal Research*, 49(1), 330–339.
- Mashatise, E., Hamudikuwanda, H., Dzama, K., Chimonyo, M. & Kanengoni, A. (2005). Effects of corn cob-based diets on the levels of nutritionally related blood, metabolites and onset of puberty in Mukota and Landrace×Mukota gilts. *Asian-Australasian Journal of Animal Science*, 18, 1469–1474.
- Maswanganye, G. M. T., Liu, B., Che, D. & Han, R. (2021). Review: Effects of dietary fiber levels and composition on the intestinal health of finishing pigs. *Open Journal of Animal Sciences*, 11, 384–398.
- McCleary, B. V., De Vries, J. W., Rader, J. I., Cohen, G., Prosky, L., Mugford, D. C., Champ, M. & Okuma, K. (2010). Determination of total dietary fiber (CODEX definition) by enzymatic–gravimetric method and liquid chromatography: Collaborative study. *Journal of the Association of Official Analytical Chemists International*, 93(1), 221–233.
- Mertens, D. R. (2003). Challenges in measuring insoluble dietary fiber. *Journal of Animal Science*, 81(12), 3233–3249.
- Meunier-Salaün, M. C., Edwards, S. A. & Robert, S. (2001). Effect of dietary fibre on the behaviour and health of the restricted fed sow. *Animal Feed Science and Technology*, 90, 53–69.
- Montagne, L., Pluske, J. R. & Hampson, D. J. (2003). A review of interactions between dietary fibre and the intestinal mucosa, and their consequences on

digestive health in young non-ruminant animals. *Animal Feed Science and Technology*, 108, 95–117.

- Mpendulo, C. T., Chimonyo, M., Ndou, S. P. & Bakare, A. G. (2018). Fiber source and inclusion level affects characteristics of excreta from growing pigs. *Asian-Australian Journal of Animal Science*, 31, 755–762.
- Nkwocha, G. A., Anukam, K. U. & Nkwocha, V. I. (2008). Performance, carcass and haematological characteristics of pigs fed graded levels of *Gmelina arborea* leaf meal. *Animal Production Research Advances*, 4(2), 148–151.
- Nwakpu, P. E., Uchewa, E. N. & Nweke, F. N. (2010). Growth performance and hematological traits of weaner pigs fed graded levels of raw bambara nut (*Vigna subterranean (L) Verdc*) waste. *Nigerian Journal* of Biotechnology, 21, 65–70.
- Oboh, S. O., Moseri, H. & Okosun, S. E. (2018). Assessment of cassava peels and palm kernel cake (pkc) on the performance of grower pig. *International Journal of Research in Agriculture and Forestry*, 5(10), 1–5.
- Obua, B. E. (2013). Haematological and biochemical profile of weaner pigs fed *Microdesmis puberula* (Hook f.) leaf meal based diets. *Animal Production Research Advances*, 9(1), 19–25.
- Ogunbode, A. A., Okeniyi, G., Fatola, O. S. G, Ogunjimi, B. A. & Folarin, M. O. (2016). Haematology and serum indices of weaner pigs fed raw pride of barbados. (*Caesalpina pulcherrima*) seed meal. *Nigerian Journal* of Animal Science, 2, 408–416.
- Oguntimein, G. B. (1992). Processing cassava for animal feed. In *Cassava as Livestock Feed in Africa*. Workshop on the Potential Utilization of Cassava as Livestock Feed in Africa, November 14<sup>th</sup>–18<sup>th</sup>, 1988, Idaban, Nigeria.
- Olayemi, W. A., Alade, A. A., Williams, G. A. & Obadae, O. O. (2006). Blood profiles, intestinal morphormetric and ceaca micro flora of broiler chickens fed with urea molasses treated maize cob. *Journal of Agricultural Science and Food Technology*, 6(5), 78–86.
- Oliviero, C., Junnikkala, S. & Peltoniemi, O. (2019). The challenge of large litters on the immune system of the sow and the piglets. *Reproduction in Domestic Animals*, 54, 12–21.
- Olukayode, A., Makinde, E. & Sonaiya, B. (2011) Utilization of sun-dried maize offal with blood meal in diets for broiler chickens. *Open Journal of Animal Science*, 1(3), 106–111.
- Paßlack, N., Vahjen, W. & Zentek, J. (2015). Dietary inulin affects the intestinal microbiota in sows and their suckling piglets. *BMC Veterinary Research*, 11, 51.

- Papatsiros, V. G., Katsarou, M. S., Drakoulis, N., Maragkakis, G., Tzika, E., Maes, D., Tassis, P. D., Lagiou, M. & Christodoulopoulos, G. (2021). Effects of dietary fibre on metabolism and performance in sows. *Polish Journal of Veterinary Sciences*, 24(2), 271–279.
- Ramonet, Y., Meunier-Salaun, M. C. & Dourmad, J. Y. (1999). High-fiber diets in pregnant sows: digestive utilization and effects on the behavior of the animals. *Journal of Animal Science*, 77, 591–599.
- Régnier, C., Bocage, B., Archimède, H. & Renaudeau, D. (2010). Effects of processing methods on the digestibility and palatability of cassava root in growing pigs. *Animal Feed Science and Technology*, 162, 135–143.
- Rose, D. J., Patterson, J. A. & Hamaker, B. R. (2010). Structural differences among alkali soluble arabinoxylans from maize (*Zea mays*), rice (*Oryza sativa*), and wheat (*Triticum aestivum*) brans influence human fecal fermentation profiles. *Journal of Agriculture and Food Chemistry*, 58, 493–499.
- Schulze, H., Van Leeuwen, P., Verstegen, M. W., Huisman, J., Souffrant, W. B. & Ahrens, F. (1994). Effect of level of dietary neutral detergent fiber on ileal apparent digestibility and ileal nitrogen losses in pigs. *Journal of Animal Science*, 72(9), 2362–2368.
- Stein, H. H. & Shurson, G. C. (2009). Board-invited review: The use and application of distillers dried grains with solubles in swine diets. *Journal of Animal Science*, 87(4), 1292–1303.
- Szyszkowska, A., Sowiński, J. & Wierzbicki, H. (2007). Changes in the chemical composition of maize cobs depending on the cultivar, effective temperature sum and farm type. *Acta Scientiarum Polonorum Agricultura*, 6, 13–22.
- Tan, C., Ji, Y., Zhao, X., Xin, Z., Li, J., Huang, S., Cui, Z., Wen, L., Liu, C., Kim, S. W., Deng, J. & Yin, Y. (2021). Effects of dietary supplementation of nucleotides from late gestation to lactation on the performance and oxidative stress status of sows and their offspring. *Animal Nutrition*, 7(1), 111–118.
- Tan, C., Wei, H., Ao, J., Long, G. & Peng, J. (2016). Inclusion of konjac flour in the gestation diet changes the gut microbiota, alleviates oxidative stress, and improves insulin sensitivity in sows. *Applied and Environmental Microbiolobiology*, 82(19), 5899–5909.
- Theil, P. K., Lauridsen, C. & Quesnel, H. (2014). Neonatal piglet survival: impact of sow nutrition around parturition on fetal glycogen deposition and production and composition of colostrum and transient milk. *Animal*, 8, 1021–1030.

- Tonukari, N. J., Oliseneku, E. E., Avwioroko, O. J., Aganbi, E., Orororo, O. C. & Anigboro, A. A. (2016). A novel pig feed formulation containing *Aspergillus niger* CSA35 pretreated-cassava peels and its effect on growth and selected biochemical parameters of pigs. *African Journal of Biotechnology*, 15(19), 776–785.
- Van der Peet-Schwering, C. M. C., Kemp, B., Binnendijk, G.
  P., Den Hartog, L. A., Spoolder, H. A. M. & Verstegen,
  M. W. A. (2003). Performance of sows fed high levels of non-starch polysaccharides during gestation and lactation over three parities. *Journal Animal Science*, 81, 2247–2258.
- Van Soest, P. J. & Wine, R. H. (1968) The determination of lignin and cellulose in acid detergent fiber with permanganate. *Journal of Association of Official Analytical Chemists*, 51, 780–785.
- Wafar, R. J., Tarimbuka, L. I., Sini, T., Adi, Z. A., Lamalang, E. B. & Bako, M. I. (2020). Growth performance and nutrient digestibility of weaner pigs fed cereal offals in diets. *Nigerian Journal of Animal Production*, 47(6), 129–134.
- Weber, T. E. & Kerr, B. J. (2012). Metabolic effects of dietary sugar beet pulp or wheat bran in growing female pigs. *Journal of Animal Science*, 90, 523–532.
- Wenk, C. (2001). The role of dietary fibre in the digestive physiology of the pig. *Animal Feed Science and Technology*, 90(1–2), 21–33.
- Woyengo, T. A., Beltranena, E. & Zijlstra, R. T. (2014). Nonruminant nutrition symposium: Controlling feed cost by including alternative ingredients into pig diets: A review. *Journal of Animal Science*, 92, 1293–305.
- Xu, C., Peng, J., Zhang, X. & Peng, J. (2020). Inclusion of soluble fiber in the gestation diet changes the gut microbiota, aects plasma propionate and odd-chain fatty acids levels and improves insulin sensitivity in sows. *Internal Journal of Molecular Sciences*, 21(2), 635.
- Yang, P. & Zhao, J. (2021). Variations on gut health and energy metabolism in pigs and humans by intake of different dietary fibers. *Food Science & Nutrition*, 9, 4639–4654.

- Yen, J. T., Nienaber, J. A., Hill, D. A. & Pond, W. G. (1991). Potential contribution of absorbed volatile fatty acids to whole-animal energy requirement in conscious swine. *Journal of Animal Science*, 69, 2001–2012.
- Yin, G., Huang, D., Zhang, H. & Wang, J. (2016). Effect of dietary fibre on serum biochemical parameters of pregnant sows. *Journal of Chemical and Pharmaceutical Research*, 6, 1222–1224.
- Zhang, S., Wu, Z., Heng, J., Song, H., Tian, M., Chen, F. & Guan, W. (2020). Combined yeast culture and organic selenium supplementation during late gestation and lactation improve preweaning piglet performance by enhancing the antioxidant capacity and milk content in nutrient-restricted sows. *Animal Nutrition*, 6(2), 160–167.
- Zhang, W., Li, D., Liu, L., Zang, J., Duan, Q., Yang, W. & Zhang, L. (2013). The effects of dietary fiber level on nutrient digestibility in growing pigs. *Journal of Animal Science and Biotechnology*, 4(1), 17.
- Zhao, J., Wang, Q., Liu, L., Chen, Y., Jin, A., Liu, G., Li, K., Li, D. & Lai, Ch. (2018). Comparative digestibility of nutrients and amino acids in high-fiber diets fed to crossbred barrows of Duroc boars crossed with Berkshire× Jiaxing and Landrace×Yorkshire. *Asian-Australasian Journal* of Animal Science, 31(5), 721–728.
- Zhuo, Y., Feng, B., Xuan, Y., Che, L., Fang, Z., Lin, Y., Xu, S., Li, J., Feng, B. & Wu, D. (2020). Inclusion of purified dietary fiber during gestation improved the reproductive performance of sows. *Journal of Animal Science and Biotechnology*, 11, 47.
- Zhou, Y., Jiang, Z., Lv, D. & Wang, T. (2009). Improved energyutilizing efficiency by enzyme preparation supplement in broiler diets with different metabolizable energy levels. *Poultry Science*, 88, 316–322.
- Ziemer, C. J., Kerr, B. J., Weber, T. E., Arcidiacono, S., Morrison, M. & Ragauskas, A. (2012). Effects of feeding fiberfermenting bacteria to pigs on nutrient digestion, fecal output, and plasma energy metabolites. *Journal of Animal Science*, 90, 4020–4027.